

ANTIBACTERIAL ACTIVITY FOR TOTAL PHENOLS AND TOTAL ALKALOIDS  
EXTRACTED FROM *NARCISSUS TAZETTA* L. BULBS

Ola Kareem Ali<sup>1\*</sup> Zainab Yaseen Mohammed Hasan<sup>2\*</sup> Arwa Abdul-kareem Tawfiq<sup>3\*</sup>  
Ghydaa H.Aljeboury<sup>4\*</sup>

<sup>1,3\*</sup> College of Science for Women, University of Baghdad, Baghdad, Iraq

<sup>2,4\*</sup> Biotechnology research center, Al-Nahrain university, Baghdad, Iraq

**Abstract**

*Narcissus tazetta* plant from Amaryllidaceae family is known to be rich in bioactive metabolites such as Alkaloids, phenolics and flavonoid which detected in almost species of this family. *N. tazetta* cultivated in Iraq, had not yet been studied for its active components antimicrobial study; thus the current study employed the antibacterial activity for total phenolic compounds and total alkaloids extracted from this plant. The plant bulb pieces were defatted with n-hexane before extraction of total phenols(TP) and total alkaloids(TA) from the plant. Results showed that the plant contains about 0.95g/140g bulb as total phenols residue and 0.7g/150g bulb as total alkaloids residue. The major Tp compounds observed in HPLC assay were Salicylic acid, Sinapic acid, vanillic acid, Caffeic acid, Chlorogenic acid, and p-coumaric acid in descending quantity. Four layer had been collected that might contain different kinds of alkaloids. The methanolic layer (1) and layer (3) which represented the chloroform layer after acidification represented the richest layers in 'Glanthamine' alkaloid represented by HPLC assay. The antibacterial as inhibitory effects of TP and TA on *E.coli*, *P.auroginosa*, *S.aureus* and *Bacillus subtilus*, showed moderate antibacterial activity for both plant extracts in coparison to some trade antibiotics.

Key words: *Narcissus tazetta*, Amaryllidaceae family, Total phenols, Total flavonoids.

Author corresponding E mail: zainaby2003@yahoo.com

**Introduction**

Medicinal properties of plants belong to Amaryllidaceae family were already known in the fourth century B.C., when Hippocrates used its oil for the treatment of uterine tumors(Miroslav et al.,2015). This family contains many plants with important biological effects among them is the plant *Narcissus tazetta* which is known worldwide for being a good source for active constituents like alkaloids and polyphenols had been used for treatment many disorders that had risen now a day as Alzheimer disease (Tomasz and Jacek ,2009), thus the plant inhibit the enzyme cholinesterase responsible for Alzheimer condition development.( Al-Snafi,2020). The plant is spring-flowering plants, and rich with biological active components; among them; cardio active glycosides, different types of flavonoids, tannins, volatile oil, beside, steroids, terpenoids, alkaloids as well as anthraquinones (Kamenetsky and Okabo ,2012). **Phenolic compounds** as a secondary metabolites are produced in almost all plant kingdom via shikimic acid pathway (Talapatra and Talapatra,2015) expressed potential in health benefits as antioxidant(Fu et al.,2016), antifungal(Hammada,2018), antiviral(Uthirapathy, 2021), anti-bacterial, cardio-protective(Chiu et al.,1992) and antitumor active agent(Ooi et al.,2000).

. In spite of their wide distribution in the plant kingdom, researchers have directed their attention to the health benefits of phenolic compounds and alkaloids (Dai and Mumper, 2010). Multi-drug resistance especially for antibiotics used today makes the infectious diseases more complicated for getting difficult to cure gave an importance towards new compounds from nature to overcome such problems. ***Phenolic compounds and alkaloids extracted from the bulbs of Narcissus tazetta cultivated in Iraq were used in the current study to investigate the antibacterial activity represent the main aim of the work.***

#### **Materials and methods:**

##### ***Plant collection***

After fresh collection of *Narcissus tazetta* bulbs from botanical garden at Baghdad city, steps for cleaning and dust removing were proceeded to prepare the plant material for subsequent experiments.

##### ***Plant total phenolic and total alkaloids Extraction***

###### ***i-Total Phenolic Extraction***

A defatting process was done before phenolic compounds extraction starting from 350 g fresh bulb pieces using n-hexan as good non-polar solvent for removing any fatty materials. The resultant weight of defatted bulb was 140 g. to be macerated with 80% ethanol and left for 7 days in cool dark place. A process of filtration and drying were done and the residue was weighted and kept in dark container for further tests (Tonisi, 2020).

###### ***ii-Extraction of Total alkaloids:*** (Balasundram et al.,2006)

About 300 gm of fresh bulbs were soaked in normal hexane for 4 days, then filtered and discarded the filtrate to get defatting plant material that allowed to dry in room temperature for solvent residual removal. The dried defatting bulbs were weighted to get 150 g, and the following steps were taken on them:

1- Soaking with 500 ml of absolute ethanol alcohol with gentle heating to a temperature of 60 degrees Celsius for an hour with stirring, the bulbs were left soaked for another 24 hours at room temperature to ensure the extraction of almost all Alkaloids in the bulb. The extract was filtered and allowed to dry and weighed, then coded as sample No. (1)

2 - The bulb residue from step 1 was re-soaked in an acidified solution with a concentration of (5% HCL) 800 ml to reach the pH (pH = 2), the soaking was left for 4 days. Then it was filtered to show a white precipitate on the filter paper, and this salt precipitate was called the sample (2). After filtration, the acid filtrate was transferred to a separating funnel, then chloroform was added to it in a ratio of 1:1, the funnel was shaken gently. The two acidified aqueous layers and the chloroform layer were separated, to produce an upper acidified aqueous layer that was used in the next step. The lower chloroform layer was dried and called the dry chloroform layer after adding the acid to the sample (3)

3- The acidic aqueous layer was taken in the previous step and a base was added with a concentration of 10% Na<sub>2</sub>CO<sub>3</sub> sodium carbonate to raise the pH value 10 to make the solution basic and then transferred to the separating funnel again and chloroform was added at a ratio of

1:1 to the solution and shaken gently. The second layer: the chloroform layer was dried. The dry layer of chloroform was named after the addition of the base sample No. (4).

All the dry layers were individually weighed and noted, and alkaloid detection was carried out using Dragendorff reagent for all layers and samples.

Samples positive for Dragendorff reagent were collected and the collected weights represent the total alkaloids present in the bulbs.

### Determination of Total Phenols and Total Alkaloids by High Performance Liquid

#### Chromatography: HPLC: (Ana-Maria et al.,2020, Özlem et al.,2019)

Table -1 showed the HPLC conditions for determination of total phenols and total alkaloids in plant bulb extracts with corresponding to standard and concentration for each

Table -1 HPLC conditions for phenols and alkaloids present in plant bulb extracts with corresponding to standard and their concentrations

HPLC Conditions	For Total phenols	For Total Alkaloids
Column	ODS L18 (10X 4.6Id) mm, 5µm particle size	ODS c18 (150*4.6 Id) mm 5mm partical site
Flow rate	0.7ml/min.	1 ml/min
Wave length /Detector	280 n.m/ uv-vis	280 n.m/ uv-vis
Temperature		35 C
Mobile Phase	A=1% Acetic acid and  B=Methanol 10%	A: 30 mmol /L Ammonium Bicarbonate + 0.7% Amonia Solution + 0.1% Tri ethyl amine  B: Acetonitnle  A/B: 30%
Standard (concentration)	vanillic acid(5µg/ml) Caffeic acid(5µg/ml) p-coumaric acid (5µg/ml) Chlorogenic acid (5µg/ml) Sinapic acid (5µg/ml) Salycilic acid(5µg/ml)	Glanthamine(6mg/ml)
Plant Extract(concentration)	12.5mg/ml	12.5mg/ml

### Antibacterial Activity for Total Phenols (TP) and Total (TA)Alkaloids :

*The determining Inhibitory Effect by well agar diffusion method of for Total Phenols and Total Alkaloids had been conducted as follow(Silva et al.,1987 ; Rayhana et al.,2020):*

A culture of *E.coli*, *P.auroginosa*, *S.aureus* and *Bacillus subtilus* previously grown in nutrient broth was streaked on Muller-Hinton agar, and then incubated under aerobic conditions at 37°C for 24 hr. After incubation a cork borer (5mm) was used to withdraw discs of each plant extract and some antibiotic disc then put on surface of the Muller-Hinton agar that was inoculated (before) with 0.1 ml of pathogenic bacteria. After incubate, at 37°C for 24 hr, the inhibition zone around the disc was estimated in (mm).

***The determination of Minimum Inhibitory Concentration (MIC) and Minimum Bactericidal Concentration (MBC) for total phenols and total alkaloids, (AL-Saadi,2016):***

The MIC test demonstrates the lowest level of antimicrobial agent that greatly inhibits growth, the MBC demonstrates the lowest level of antimicrobial agent resulting in kill microorganism. Different concentrations of each plant extract were made in tubes containing sterile nutrient broth each. The concentrations were (1/9, 2/8, 3/7, 4/6, 5/5, 6/4, 7/3, 8/2 and 9/1) giving final volume 10ml in each tubes. Then each concentration was inoculated by 0.1ml culture previously grown in nutrient agar *E.coli*, *P.auroginosa*, *S.aureus* and incubated at 37 °C for 24 hr. After incubation the growth of tubes was observed and minimum inhibitory concentration and was determined as the lower concentration of the filtrate that gave no growth of each bacteria in the tubes, and Minimum Bactericidal Concentration MBC demonstrates the lowest level of antimicrobial agent resulting in kill pathogenic bacteria.

## **Results and Discusion**

### **Plant total phenolic and total alkaloids yields from Extraction**

The total phenol residue weight was 0.9544g/140g defatted bulbs. While the residue for total alkaloids extracted from bulbs were estimated by the summation of the weight of all layers gave positive results with dragendroff's test and that equal to about 0.7 g/150g defatted bulbs.

### **Determination of Total Phenols and Total Alkaloids by High Performance Liquid Chromatography: HPLC:**

Figure (1) and (2) investigated the major phenolic compounds in the standard and sample extracted solution respectively. Table (2) showed the major phenolic compounds and their concentration detected in the plant bulb.

Figure (3) and (4) represented the HPLC chromatogram for the standard alkaloid 'Glanthamin' and the defatted bulbs.

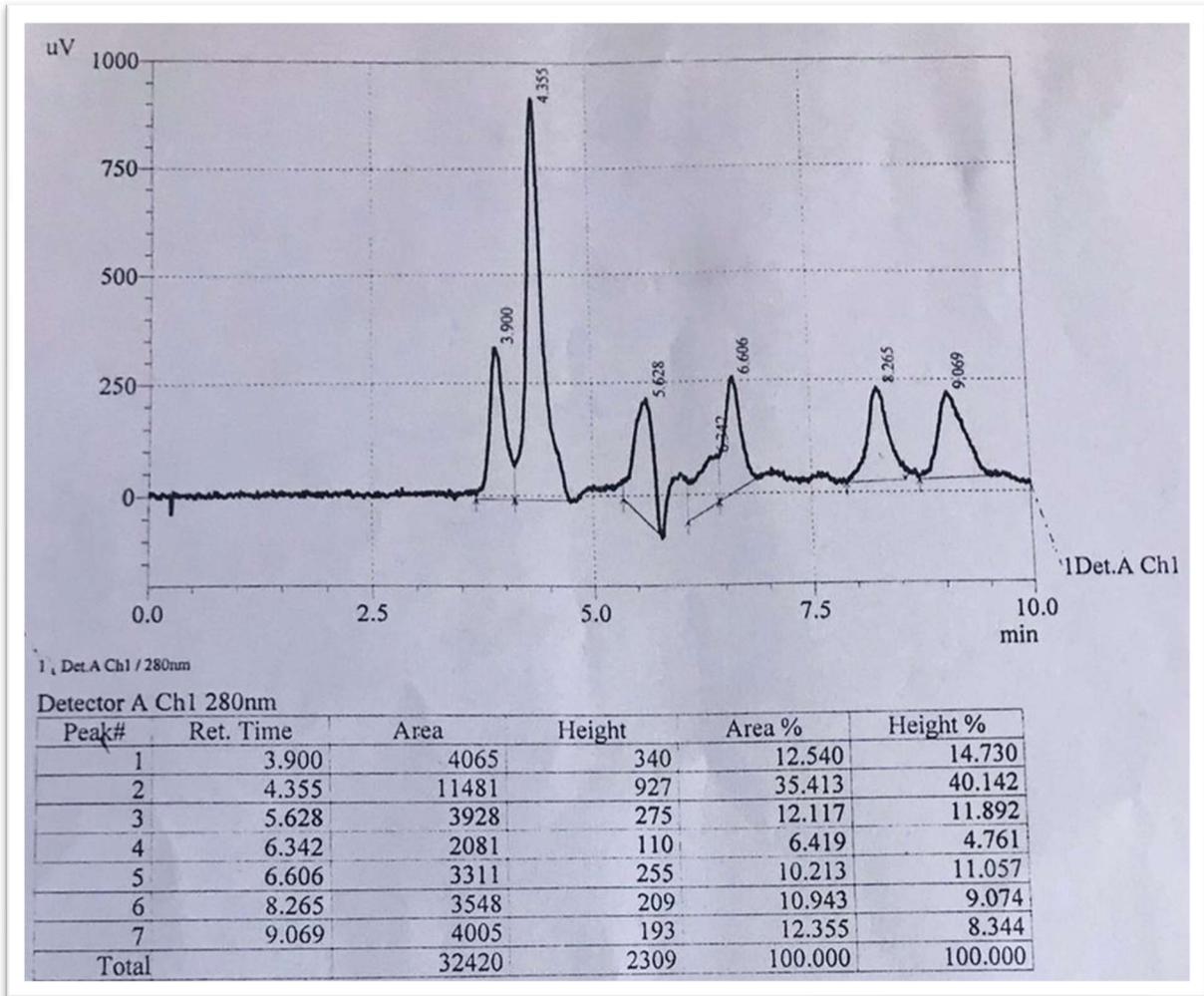


Figure (1) HPLC chromatogram for standard phenolic compounds

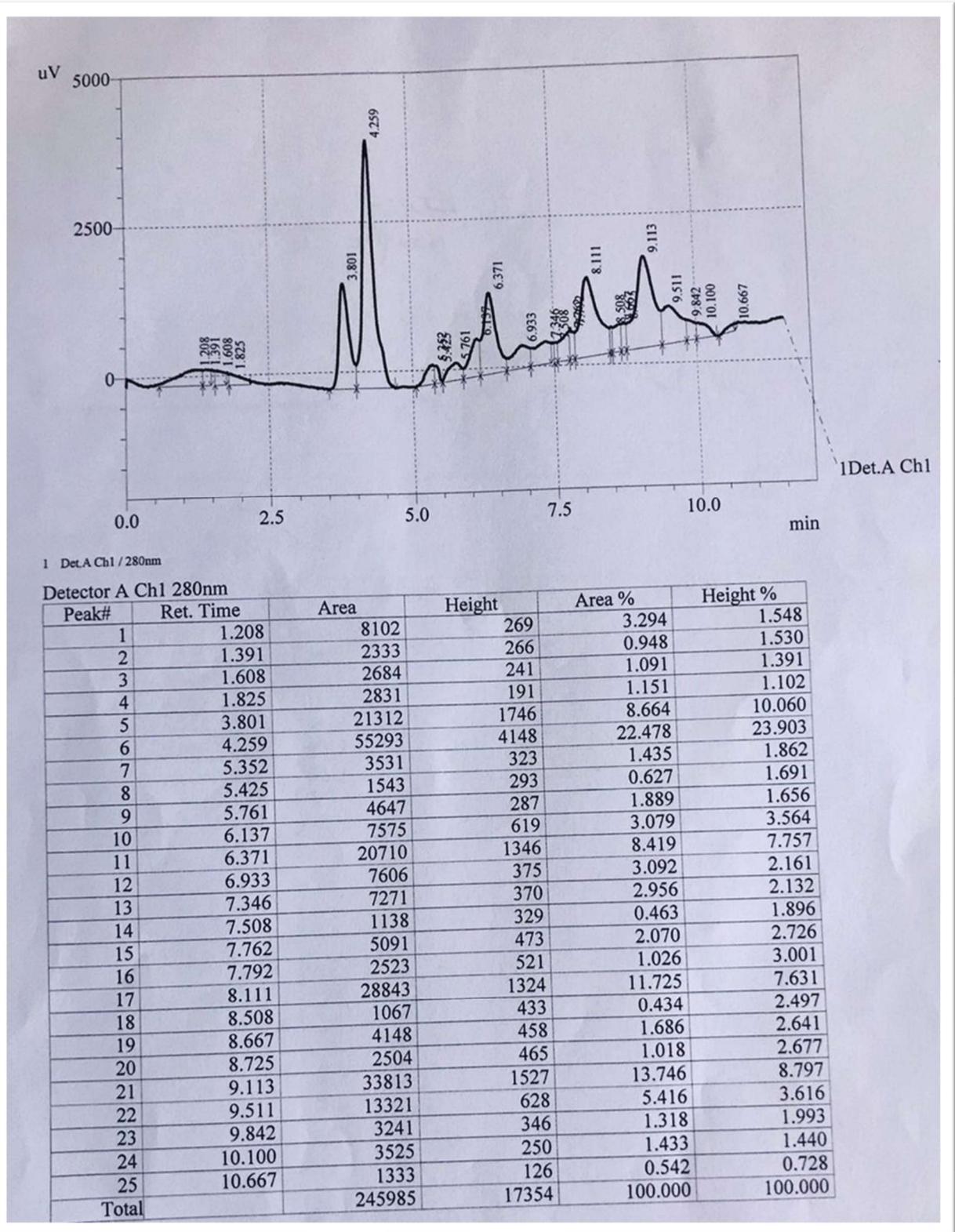


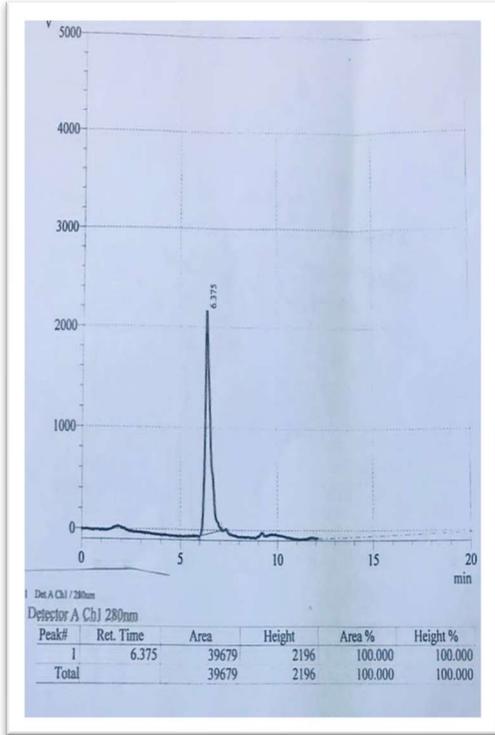
Figure (2) HPLC chromatogram for plant extracted phenolic compounds

Table (2) HPLC analysis results for standards and the extracted phenolic compounds

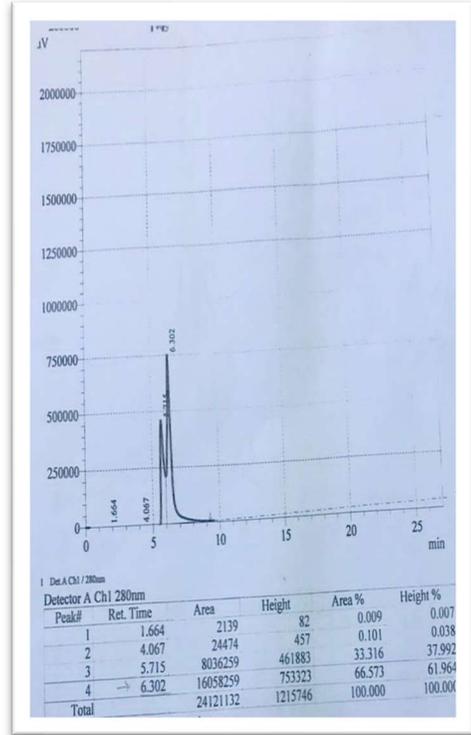
Phenolic compound	Conc. $\mu\text{g} / \text{ml}$	Rt.in minutes For Standarad phenols	Rt.in minutes For the extracted phenols	Concentration $\mu\text{g} / \text{g}$ plant
vanillic acid	5	3.900	3.801	26.2
Caffeic acid	5	4.355	4.259	24
p-coumaric acid	5	5.628	5.761	6
Chlorogenic acid	5	6.606	6.933	11.5
Sinapic acid	5	8.265	8.111	40.6
Salycilic acid	5	9.069	9.113	42.2

The plant bulb seemed to be good source of several phenolic compound, the most abundant was salycilic acid(42.2  $\mu\text{g} / \text{g}$  ) and sinapic acid(40.6  $\mu\text{g} / \text{g}$  ), with less extend vanillic acid (26.2  $\mu\text{g}/\text{g}$ ), caffeic acid(24  $\mu\text{g} / \text{g}$ ), chloragenic acid (11.5  $\mu\text{g}/\text{g}$ ) and the least content was p-coumaric acid(6  $\mu\text{g}/\text{g}$ ).

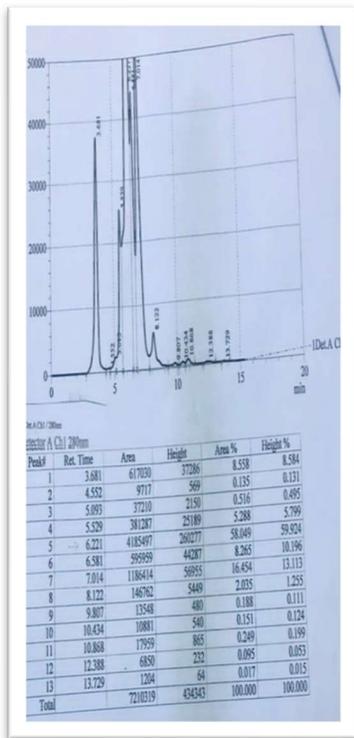
The HPLC chromatogram for alkaloids in the four extracted layer from plant bulbs were represented in the figures(3) and table (3) in respect with standard alkaloid "Glanthamine". A few studies on the natural compounds present in the narcissus plant grown in Iraq and its biological activity against microbes, infections or various diseases had been conducted (Bushra and Mahmood, 2018). The phytochemical constituents and their concentration for this plant was employed in the current study differ from what is found in *Bati* and co. study (Bati et al.,2018)



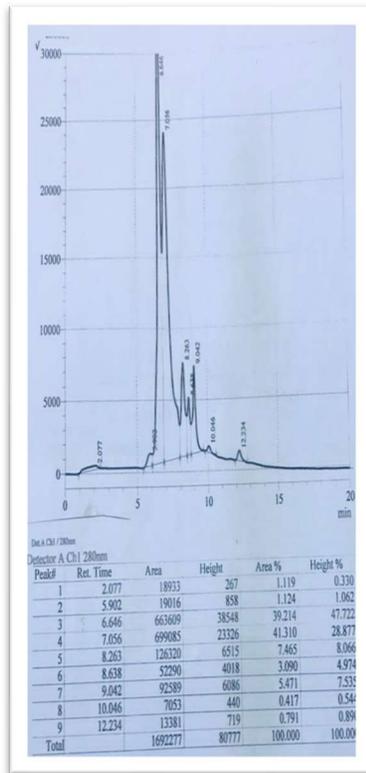
**Glanthamine standard alkaloid**



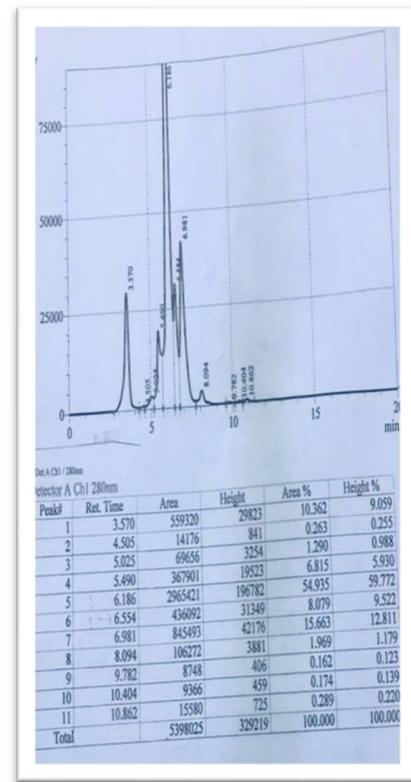
**Layer 1**



**Layer 2**



**Layer 3**



**Layer 4**

Figure (3) HPLC chromatogram for Glanthamine alkaloid standard and plant extracted Alkaloids in four layers (1,2,3,4)

Table (3) HPLC analysis results for standards and the extracted Alkaloids in four layers

Alkaloid / Layer	Area under the curve	Rt.in minutes	Concentration "Glanthamine" mg / ml
<b>Glanthamine standard</b>	<b>39679</b>	<b>6.375</b>	<b>0.006</b>
<b>Layer1</b>	<b>16058259</b>	<b>6.302</b>	<b>2.43</b>
<b>Layer2</b>	<b>Not found</b>	<b>-----</b>	<b>-----</b>
<b>Layer3</b>	<b>4185497</b>	<b>6.221</b>	<b>0.633</b>
<b>Layer4</b>	<b>436092</b>	<b>6.554</b>	<b>0.066</b>

Three layers appeared to contain Glanthamine alkaloid. Layers 1 and 3 were superior in containing the largest amount of the alkaloid mentioned

#### Antibacterial activity for Total Phenols and Total Alkaloids:

Determining inhibitory effects of TP and TA on *E.coli*, *P.auroginosa*, *S.aureus* and *Bacillus subtilus* in comparison to trade antibiotic by well agar diffusion method were shown in table(4), while figures(4),(5),(6), and (7), declare the zones shapes and diameters represented the inhibitory effects of TP,TA, and each extracted alkaloid layer toward the four pathogenic bacteria used in this study. In spite that layer 2 contained trace amounts of Alkaloid as shown in Dragendroff test or almost free from alkaloids as not detected by HPLC assay, this layer was employed for antibacterial assay.

Table(4)The inhibitory effect as inhibition zone of TP and TA on the four pathogenic bacteria in comparison to some antibiotic and their concentrations

Plant Extract or Antibiotic disc	Conc. (mg/ml)	Inhibition Zones (mm)			
		<i>E.coli</i>	<i>P.auroginosa</i>	<i>S.aureus</i>	<i>Bacillus subtilus</i>
Total phenol(TP)	8	17	17	21	18
Total Alkaloid(TA)	8	16	20	23	22
Alkaloid Layer 1	8	13	11	12	13
<b>Alkaloid Layer 2</b>	<b>8</b>	<b>11</b>	<b>16</b>	<b>13</b>	<b>14</b>
Alkaloid Layer 3	8	12	12	15	14
Alkaloid Layer 4	8	19	11	14	25
TE-antibiotic	0.03	10	10	12	13
CTX-antibiotic	0.03	13	13	15	15
AX-antibiotic	0.025	10	13	13	13

IPM-antibiotic	0.01	20	20	15	20
AK-antibiotic	0.01	9	15	15	15
CIP-antibiotic	0.01	25	26	30	29

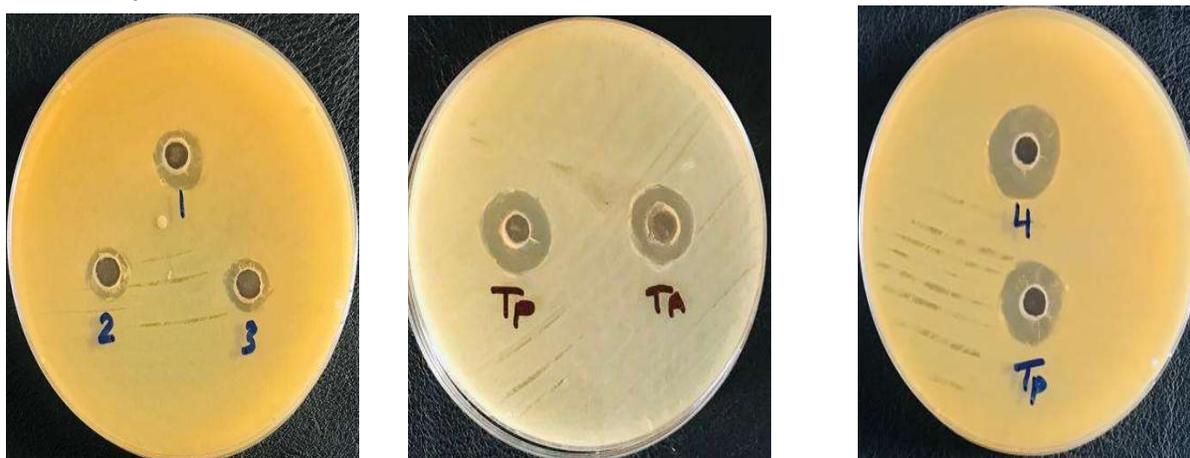
Since the discovery of penicillin, researchers conducted their affords toward finding other natural products with promising to be new drug candidates in treating antibiotic-resistant infections specially in multi resistance pathogens(Chee et al.,2021, Benedec et al.,2018).

Phenolic Compounds extracted from different plants play important role in killing and inhibiting the harmful tinny microorganism as these natural substance known to have antioxidant potency due to the hydroxyl group in their structure.(Aziz et al.,2022),

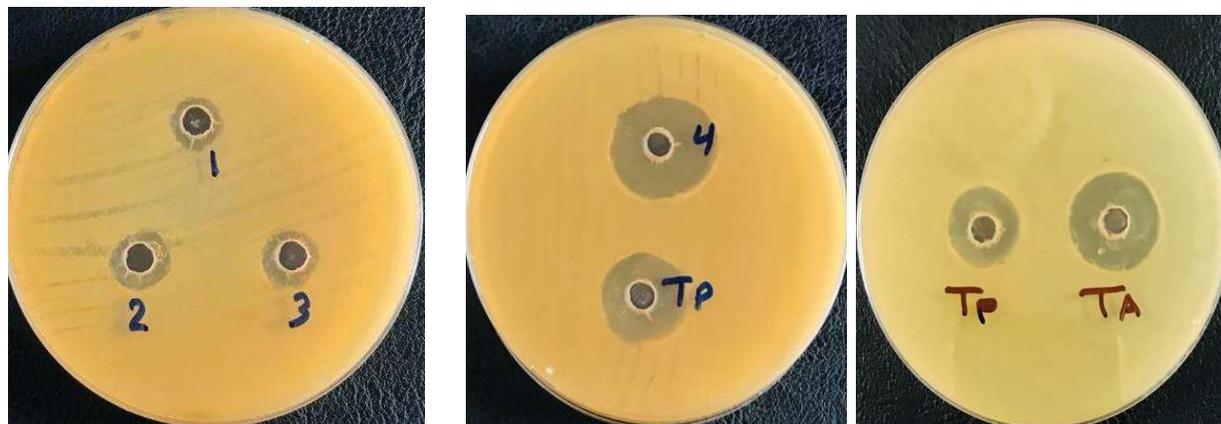
In general; total alkaloids (TA) appeared to be more potent than Total phenolic(TP) compounds extracted from plant bulbs. The different types of alkaloids extracted in each layer (except layer 2) affected the pathogenic bacteria in different manner. Table (4) declared that layer 4 possessed the potent antimicrobial effects as the of the antibiotic "imperamine"(IPM) activity.



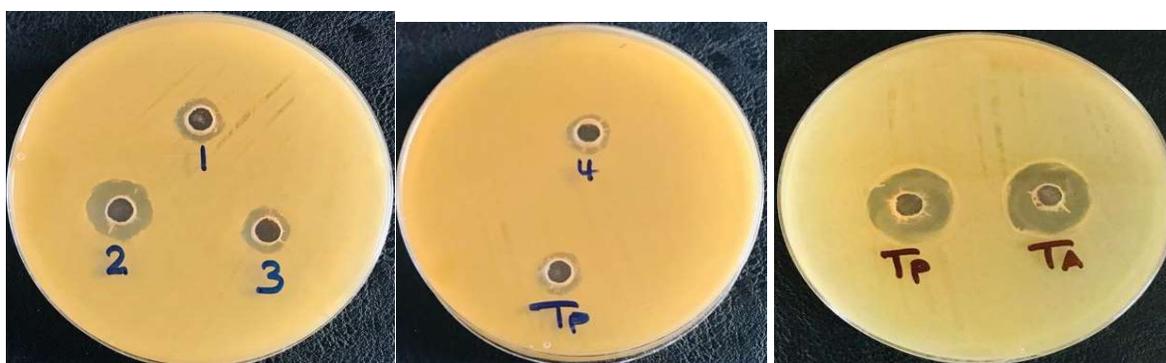
Figure(4) The inhibitory zones diameters against *Staphylococcus.aureus* byTP,and TAwith four extracted layers



Figure(5) The inhibitory zones diameters against *Escherichia coli* by TP,and TA with four extracted layers



Figure(6) The inhibitory zones diameters against *Bacillus subtilis* by TP, and TA with four extracted layers



Figure(7) The inhibitory zones diameters against *P. auroginosa* by TP, and TA with four extracted layers

For the determination of Minimum Inhibitory Concentration (MIC) and Minimum Bactericidal Concentration (MBC) of the total phenols (TP) and total alkaloid(TA) for the plant extract, table(4) and(5) investigate the MIC and MBC value respectively for the three pathogenic bacteria used in this study

Table (5)The Minimum Inhibitory Concentration for TP and TA Against *Staphylococcus.aureus*, *Escherichia coli* and *P.auroginosa*

Plant Extract	MIC value( $\mu\text{g/ml}$ ) against <i>S.aureus</i>	MIC value( $\mu\text{g/ml}$ ) against <i>E.coli</i>	MIC value( $\mu\text{g/ml}$ ) against <i>P.auroginosa</i>
TP	5.04	5.04	4.032
TA	4.032	4.032	3.6288

Table (6) The Minimum Bactericidal Concentration for TP and TA Against *Staphylococcus aureus*, *Escherichia coli* and *P.auroginosa*

Plant Extract	MBC value( $\mu\text{g/ml}$ ) against <i>S.aureus</i>	MBC value( $\mu\text{g/ml}$ ) against <i>E.coli</i>	MBC value( $\mu\text{g/ml}$ ) against <i>P.auroginosa</i>
TP	7.2	7.2	5.04
TA	5.04	5.04	4.032

The inhibitory concentration (MIC) for total phenols (TP) extracted from the bulbs against *S.aureus* and *E.coli* represented by the concentration of 1.44  $\mu\text{g/ml}$ . while for *P.auroginosa* bacteria MIC appeared at 0.288  $\mu\text{g/ml}$ . For the bactericidal minimum concentration (MBC) were 4.8  $\mu\text{g/ml}$  for *S.aureus* and *E.coli* and 1.44  $\mu\text{g/ml}$  *P.auroginosa* bacteria. In case of total Alkaloids (TA), The MIC against *S.aureus* and *E.coli* showed at 0.288  $\mu\text{g/ml}$  which was more potent effect than TP even against *P.auroginosa* the minimum inhibitory concentration was 0.0288  $\mu\text{g/ml}$ . The total alkaloid (TA) MBC against *S.aureus* and *E.coli* was 1.44  $\mu\text{g/ml}$  and 0.288  $\mu\text{g/ml}$  against *P.auroginosa*.

#### Conclusion:

*Narcissus tazetta* bulbs cultivated in Iraq considered as good source for phenolic compounds either as simple phenolic or flavonoids, besides their alkaloids rich components. The major TP compounds observed in HPLC assay were Salicylic acid, Sinapic acid, vanillic acid, and Caffeic acid. Different alkaloids had been extracted and the famous is the Glanthamine alkaloid which was extracted in the acidified chloroform layer represented the richest layers in this alkaloid by HPLC assay. The antibacterial effects of TP and TA against *E.coli*, *P.auroginosa*, *S.aureus* and *Bacillus subtilis*, showed moderate antibacterial activity for both plant extracts in comparison to some trade antibiotics, that will encourage researchers to high light and conduct more efforts about investigating the plant active compounds of other medical or even prophylaxis effect.

#### Acknowledgment

*Authors would like to acknowledge the collage of Science for women / University of Baghdad and Biotechnology Research Center/Al-Nahrain University/ Ministry of Higher Education for support in completing all work requirements.*

#### References

- (1) Miroslav, L.; Jitka, N.; Pavel, K.; Anna, H.; Ladislav, K.; Lucie, G.; Marcela, Š.; Lubomír, O. and Lucie, C. (2015). Antifungal and Antibacterial Activity of Extracts and Alkaloids of Selected Amaryllidaceae Species *Natural Product Communications* Vol. 10 (9).
- (2) Tomasz, M. and Jacek, M. (2009). Pressurized liquid extraction and anticholinesterase activity-based thin-layer chromatography with bioautography of *Amaryllidaceae* alkaloids. *Analytica Chimica Acta* 633, 188–196

- (3) Al-Snafi, A.E. (2020). Constituents and pharmacology of *Narcissus tazetta*. IOSR Journal of Pharmacy. 10(9): 44-53.
- (4) Kamenetsky, R; Okubo, H. (2012). Ornamental Geophytes: From Basic Science to Sustainable Production; CRC Press: Boca Raton, FL, USA.
- (5) Talapatra, S. K. and B. Talapatra. (2015). Shikimic acid pathway. pp. 625–678. Chemistry of Plant Natural Products: Stereochemistry, Conformation, Synthesis, Biology and Medicine. Springer Berlin Heidelberg, Berlin, Heidelberg.
- (6) Fu, K.L.; Li, X.; Ye, J.; Lu, L.; Xu, X.K.; Li, H.L.; Zhang, W.D. and Shen, Y.H. (2016). Chemical constituents of *Narcissus tazetta* var. *chinensis* and their antioxidant activities. *Fitoterapia* 113: 110-116.
- (7) Hammada, H.M. (2018). Phytochemical and Biological Investigation of *Narcissus pseudonarcissus* Cultivated in Egypt. *Rec. Pharm. Biomed. Rec. Pharm. Biomed. Sci.* 2(1): 26 -34.
- (8) D. S. Uthirapathy, “Cardioprotection effects of diosgenin from *Dioscorea bulbifera* against isoproterenol-induced myocardial infarction,” *Drugs and Cell Therapies in Hematology*, vol. 10, no. 1, pp. 887–896, 2021.
- (9) Chiu, K.W.; Lee, Y.C. and Yung, K.H. (1992). Bioactive substances from the Chinese daffodil, *Narcissus tazetta*. *Phytotherapy Research* 6(5): 231-236
- (10) Ooi, L.S.; Ng, T.B.; Geng, Y. and Ooi, V.E.. (2000). Lectins from bulbs of the Chinese daffodil *Narcissus tazetta* (family Amaryllidaceae). *Biochem Cell Biol*; 78(4): 463-468.
- (11) Dai, J. and R. J. Mumper. (2010). Plant phenolics: Extraction, analysis and their antioxidant and anticancer properties. *Molecules* 15: 7313–7352.
- (12) Tonisi, S., K., L. (2020). Evaluation of bioactive compounds, free radical scavenging and anticancer activities of bulb extracts of *Boophone disticha* from Eastern Cape Province, South Africa. *Saudi Journal of Biological Sciences* 27: 3559–3569.
- (13) Balasundram, N. Sundram, K. and Samman, S. (2006). Phenolic compounds in plants and agri-industrial by-products: Antioxidant activity, occurrence and potential uses. *Food Chem.* 99: 191–203.
- (14) Ana-Maria, S.; Ion, T. and Sina, C. (2020). Determination of Phenolic Compounds Using HPLC-UV Method in Wild Fruit Species. *Horticulturae* 2022, 8, 84
- (15) Özlem, B. ;ACIKAR, A.; Betül, S. ;YILMAZ, D.; YAZGAN, G.; İŞCAN, S. (2019). Quantification of Galantamine in *Sternbergia* Species by High Performance Liquid Chromatography *Turk J Pharm Sci* 2019;16(1):32-36.
- 16 )Silva, M.; Jacobus, N. V.; Deneke, C. and Gorbach, S. L. (1987). Antibacterial substances from a human *Lactobacillus* strain. *Antimicrob. Agent and Chemother.*; 31 (8): 1231-1233.
- (17). Rayhana, S.N.; Zainab, Y.M. H. and Rawaa AlChalabi. (2020). Study the antimicrobial activity of ethanolic extract of *Lepidium draba* on some skin infectious agents. *Journal of Biotechnology Research Center* Vol. 14 No.1 pp 10-20.

- (18)AL-Saadi,Z.N. (2016). Estimation of minimum inhibitory concentration (MIC) and minimum bactericidal concentration (MBC) of cell-free extracts of Bifidobacterium species against methicillin resistant Staphylococcus aureus in vitro, Am. J. Biomed. Life Sci. vol.4 page 75-80.
- (19) Bushra, M. A. and Mahmood, O. A. (2018). Evaluating the Genotoxicity Enhancement of the Antileukemic Drug 6-Mercaptopurin When Combined With Iraqi *Nerium oleander* and *Narcissus tazetta* Extracts in vivo. Journal of Biotechnology Research Center Vol. 12 No.1 Pp92-102
- (20) Bati, A. E.; Gül, M.; ;Açikgöz, A.; Yarılgaç, T.; Kara, Ş. M. (2018) Assessment of Antioxidant Activity of Giant Snowdrop (*Galanthus elwesii* Hook) Extracts with Their Total Phenol and Flavonoid Contents. Indian Journal of Pharmaceutical Education and Research 52 (4):128-132.
- (21) Chee, K.K.; Liang, E.L.; Wei, S. S.; Wei-Hsum Y.; Kooi-Yeong K.; Long, C. M.; Andrei, M.; Bey-H. G. and Poh, H.(2021). Biological Activities of Snowdrop (*Galanthus* spp., Family Amaryllidaceae) Frontiers in Pharmacology, Volume 11 | Article 552453.
- (22). Benedec, D.; Oniga, I.; Hanganu, D.; Gheldiu, A.M.; Puşcaş, C.; Silaghi-Dumitrescu, R.; Duma, M.; Tiperciuc, B.; Vârban, R.; Vlase, L. (2018) Sources for developing new medicinal products: biochemical investigations on alcoholic extracts obtained from aerial parts of some Romanian Amaryllidaceae species. BMC Complementary and Alternative Medicine 18 (226): 1-12
- (23) Aziz, B. ; Fouzia, R.F.; Yassine, O.; Mohamed, O.; Rahou, A.; Emilia, C.; Natalizia, M.;Maria, F.T.; Luigi, M. and Francesco, C. (2022).Phenolic Compounds, Antioxidant and Antibacterial Activities of Extracts from Aerial Parts of *Thymus zygis* subsp. *gracilis*, *Mentha suaveolens* and *Sideritis incana* from Morocco .Chem. Biodiversity,vol. 19,1-13.